A systematic study of opisthorchiasis prevalence in Russia started as early as the 1920s. Over 40 years, the All-Union Helminthological Expeditions, initiated and supported by Academician K.I. Skryabin, have explored almost all area of the Soviet Union. Thus, they found that the distribution range of the main opisthorchiasis agent, *Opisthorchis felineus* (feline or Siberian fluke), in this country is vast and spans from the Biryusa River in the east to the western borders of the country.

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Currently, in addition to Western Siberia, liver flukes are found in the Volga–Kama, Don, and Dnepr River basins and of the adjacent countries, in Kazakhstan, Belarus, Ukraine, and Baltic states. Map shows the sampling sites for the liver flukes deposited in the collection of the Institute of Cytology and Genetics, Siberian Branch, Russian Academy of Sciences (Novosibirsk). She is the author and coauthor of over 100 research papers and five author’s certificates and patents.
The first intermediate host of the parasitic trematodes belonging to the family Opisthorchiidae is the freshwater snails *Bithynia*. Within the snails, trematode larvae emerge from eggs to undergo several stages of metamorphosis eventually becoming cercariae. These motile larvae leave their host for aquatic medium, where they find and enter their second intermediate host, freshwater carp fish. In the fish muscles, cercariae encyst to give metacercariae. Eating such infected fish, the final host of this parasite, mammals and, in particular, humans, are infested too (in the case of experimental infection, laboratory animals – golden hamsters – are fed the metacercariae isolated from infested fish).

In the mammalian liver and bile ducts, metacercariae metamorphose into *Opisthorchis felineus*, the major opisthorchiasis agent in this country, and about 300 specimens of other Opisthorchiidae species, including those of epidemiological importance. These studies, purely theoretical from the first glance, have an important applied perspective, since the genetic differences between parasites are the particular factors that determine significant medical characteristics, such as diversity in clinical manifestations, probability for emergence of drug resistance, and different immunogenicity (the ability to induce production of specific antibodies) of the parasites. All these data are important for prediction of the disease course and induction of the immune response in infested individuals as well as for elaboration of diagnostic tools and vaccines.

Since 2005, the Institute of Cytology and Genetics (Novosibirsk) is involved in such an integrated research into opisthorchiasis and its pathogens. Analysis of the population genetic structure of these helminths started from creation of a unique collection comprising the liver flukes from different geographic localities, namely, Western Siberia, European Russia, and Northern Kazakhstan. So far, this collection contains about 300 specimens of *Opisthorchis felineus*, the major opisthorchiasis agent in this country, and about 300 specimens of other Opisthorchiidae species, including those of epidemiological importance.

Method as a basis

The genetic diversity of helminths from different habitats has been assessed by comparative analysis of genomic markers, namely, DNA fragments. This method, widely used in current molecular biology, allows for a reliable estimation of the degree of kinship for both individuals and their groups. Therefore, it is used for determining closely related species and reconstructing their evolutionary history.

Typically, the single nucleotide polymorphisms (that is, single “letters” of the genetic code differing in the compared DNA sequences) in a certain selected DNA region rather than the whole region are used as such markers. The rate of such single nucleotide substitutions (mutations) that appear due to random reasons is very low, amounting to approximately $10^{-8}–10^{-12}$ per cell per generation. That is why the genome regions with a high rate of mutation accumulation are selected as genomic markers. This is characteristic of the genome regions that for some reasons or other escape the natural selection, that is, are selectively neutral.

The nuclear DNA sequences not coding for any proteins or the DNA of cell organelles, mitochondria, meet this requirement. The latter is inherited in a strictly maternal manner with the egg cell cytoplasm and is not involved in recombination (exchange of chromosome regions during meiosis).
Genetic similarity between different geographically isolated *O. felineus* populations was assessed using two mitochondrial genetic markers: (a) cox1 and (b) ITS1. As has been shown, this species is rather genetically uniform over the entire distribution area: over 70% of the sets of single nucleotide polymorphisms (in figure, central circle) are present in all examined populations without any exception. The remaining polymorphisms (in figure, small circles) account for less than one-third and may occur in individual populations.

The accumulation rate for neutral mutations in mitochondrial genes is five–tenfold higher as compared with the genes encoded in the nuclear DNA. It is commonly believed that the number of accumulated single nucleotide substitutions in selectively neutral DNA regions displays linear time dependence. Correspondingly, a mere counting of these substitutions may give an approximate dating of evolutionary events.

In order to find species-specific genome markers, it is first necessary to sequence a genome (that is, to “read” its nucleotide sequence). Correspondingly, the initial research task was to sequence some fragments of the nuclear DNA and the complete mitochondrial genome of *O. felineus*, which was successfully done at the Institute of Cytology and Genetics under an integrated project on liver flukes. Thus, eight suitable genetic markers were detected and three of them (two mitochondrial and one nuclear) were selected for genotyping the collection specimens of liver flukes. Note also that although analogous markers have been recently used in population and phylogeographic studies of other parasitic trematodes in Europe, East and Southeast Asia, Africa, and America, *O. felineus* was left beyond this research.

A “bottleneck” of the Ice Age

The genome markers designed at the Institute of Cytology and Genetics have allowed for “sorting out of the mess” in the family of liver flukes, which, in particular, contains two closely related species—the above mentioned *O. felineus* and *Clonorchis sinensis*, causing clonorchiasis, the disease very similar to opisthorchiasis. The analysis demonstrates that these two species diverged from a common ancestor approximately three million years ago.

The subsequent evolutionary history of *O. felineus* seems rather dramatic. The fact is that three liver fluke subspecies were earlier hypothesized, namely, Siberian, Kazakhstan, and Eastern European, correspondingly inhabiting the
Ob–Irtysk and Yenisei, Nura–Sarysuk, and Volga, Don, and Ural River basins (Beer, 2005). Therefore, it was expected that the collection specimens from so geographically distant populations would display noticeable genetic distinctions, since the ecological conditions were also drastically different. However, despite these assumptions, the examined liver fluke specimens displayed a very low genetic diversity unlike the earlier studied trematode species (Brusentsov et al., unpublished data).

What is the reason of such an amazing genetic uniformity of liver flukes? Presumably, this uniformity is a result of drastic reduction in the size of the only ancestral *O. felineus* population in Eurasia that survived after the Pleistocene glaciations. These glaciations were accompanied by a reduction in the habitats of liver fluke intermediate hosts (freshwater mollusks and fish) as well as in their infestation rates, which at that time did not exceed 2 % (Beer, 2005). This phenomenon—when population passes through the stage of a critically low size— is referred to as the “bottleneck”; as a rule, a bottleneck results in a drastic impoverishment of the population gene pool.

With further warming and development of new river basins, liver flukes restored the size of their ancestral population. According to the estimations, the period of population explosion in the history of this species started 21—25 thousand years ago (and continues until now); presumably, a facultative hermaphroditic reproduction of the liver fluke significantly enhanced this explosion.

Note that the above mentioned trematode species other than *O. felineus*, which are also able to reproduce in a hermaphroditic manner, still display considerably higher genetic diversity. However, a relatively “poor” *O. felineus* gene pool has not prevented this species from restoring and successfully expanding its distribution range.

Unfortunately, the conducted study does not allow the location of ancestral liver fluke population to be precisely determined and its dispersal routes to be traced. Nonetheless, taking into account an intricate *O. felineus* life cycle, it looks quite evident that the direction and rate of its expansion have been determined by the migratory capacities of its hosts. The fact that opisthorchiasis is currently prevalent only in certain geographic localities is in many respects determined by the environmental preferences of *O. felineus* first intermediate host—the freshwater mollusk *Bithynia*, which dies in seawater. However, human, having almost no geographic barriers, is the definitive host of liver flukes.

**In one tube**

Genetic markers also have allowed a topical medical problem of a precise species identification of liver flukes to be solved. The fact is that along with *O. felineus*, another epidemiologically significant species, *Metorchis bursa*, circulates in Russia, Kazakhstan, and Western European countries. As for the Far East, additional trematode species *C. sinensis*, common for Southeast Asian countries, is found there.

All these trematodes cause diseases with very similar clinical manifestations, interfering with precise diagnosis based on the disease symptoms alone. Microscopic examination of the eggs of these helminths also fails to distinguish between the species due to their similarity; so that the diagnosis based on testing the fecal or duodenal samples depends on the qualification of laboratory assistant. The situation is aggravated by the possibility of mixed infection. Today, PCR diagnostics of the diseases caused by trematodes gives the most precise results. PCR-based test kits have been designed abroad since the 1990s. In this country, the start was delayed; however, PCR diagnostic kits for opisthorchiasis have been actively elaborated in Russia, in particular, at the Institute of Cytology and Genetics. These diagnostic kits are able to detect genetic fragments strictly specific of a particular pathogen species in any laboratory samples. Since this method allows genetic markers for several helminths to be simultaneously used, one test is sufficient for a precise identification of parasitic agents.

Species identification of a parasite is of great importance. For example, the infection source and site can be thus determined. But the most important thing is that the agents of opisthorchiasis, metorchiasis, and clonorchiasis differ in a number of biological characteristics, which may influence the disease course and prognosis, potential complications, and the degree of parasite’s sensitivity to drugs.

In particular, it has been experimentally demonstrated in golden hamster model that two liver fluke species, *O. felineus* and *O. viverrini*, both met in Southeast Asian countries, differ in the degree of their attack on the host organism (*Lvova* et al., 2012). By the way, another trematode similar to *O. viverrini*, *Haplorchis taichui*, is widespread in golden hamster model that two liver fluke species, *O. felineus* and *O. viverrini*, both met in Southeast Asian countries, differ in the degree of their attack on the host organism (*Lvova* et al., 2012). By the way, another trematode similar to *O. viverrini*, *Haplorchis taichui*, is widespread.
in Southeast Asia; however, infection with this trematode does not lead to a severe disease (Lovis et al., 2009). This example shows how important is to differentially diagnose trematodiases for an adequate treatment prescription.

In addition, a chronic trematode invasion can lead to serious health complications even at a low infection rate. For instance, infection with this trematode in Southeast Asia; however, infection with this trematode does not lead to a severe disease (Lovis et al., 2009). This example shows how important is to differentially diagnose trematodiases for an adequate treatment prescription.

Currently, the most widespread is serological diagnostics, in particular, enzyme immunoassay (EIA), and a number of test kits are commercially produced for this purpose. This approach is based on detection of the antibodies specific to antigens of the parasite in the patient’s blood serum. A flaw of EIA is its poor sensitivity and specificity: it rather fails when the antibody concentration is low and may give a false positive result if the diagnostic antigen reacts with a molecular target other than the “necessary” antibodies.

The accuracy of EIA may be increased by cloning the genes encoding the antigens specific of the liver fluke. Such genetically engineered antigenic proteins of *O. felineus* suitable for designing new generation diagnostic test kits have been obtained at the Institute of Cytology and Genetics. They may be used for detection of most minute amounts of specific antibodies to the parasite in the blood serum. However, it takes time for these diagnostic kits to be widely used in medical practice.

Undoubtedly, introduction of novel medical technologies will influence the treatment quality and reduce the rehabilitation period. There are many examples of this kind, and most illustrative of them are associated with diagnostics of bacterial infections, which passed from petri dishes to PCR tubes. However, the novel technologies do not annul the traditional assays but rather considerably increase the sensitivity and accuracy of tests, especially for complex clinical cases.

The test kits for PCR-based diagnostics of opisthorchiasis designed by Novosibirsk scientists have successfully passed laboratory trials, and their testing with clinical samples is close to completion. These diagnostic kits already allowed mixed trematode invasions to be detected in a group of patients (Brusentso et al., 2010). This evidently demonstrates that such diagnostic tests should be included into the toolkit for laboratory diagnostics of opisthorchiasis. Production technology for these diagnostic kits is under development.

Nonetheless, the DNA-based diagnostics for helminthiases have not yet become a routine medical practice.

Today, the most precise method in diagnosing trematodiases is PCR-based approach. The test kits for differential PCR* diagnostics of opisthorchiasis agents based on several genetic markers have been developed at the Institute of Cytology and Genetics, Siberian Branch, Russian Academy of Sciences. 

Left, electrochemical pattern (agarose gel) of the PCR products amplified from the DNA of four epidemiologically significant liver fluke species, namely, *O. felineus*, *O. viverrini*, *C. sinensis*, and *M. bilis* (molecular weights of fragments are shown).

* PCR: polymerase chain reaction, is a method for synthesizing a large number of copies of an initial DNA fragment.

The knowledge about molecular genetic diversity of this parasite yielded by this project will further project the background for detecting the therapeutic targets for opisthorchiasis treatment, which is no less important than its precise diagnosis. Note that only drug, praziquantel, almost exclusively used for the chemotherapy of opisthorchiasis, has several negative side effects.

On the other hand, we should not underestimate the significance of this new highly specific and sensitive method allowing for a precise identification of the pathogen and correspondingly, for prescription of timely and adequate treatment. The method currently used for diagnostics of opisthorchiasis (light microscopy assay of fecal samples for helminth eggs) requires highly qualified personnel, which is rather in demand. DNA diagnostics requires more complex equipment but considerably decreases the human factor aspect. A wide use of DNA diagnostics requires DNA isolation from clinical material (fecal samples), and this problem was also solved under this project.

Indeed, EIA allows for detection of the antibodies to the liver fluke. However, these two methods, PCR and EIA, should not be opposed but rather regarded as counterparts of the diagnostic process. EIA is more appropriate for controlling the treatment via monitoring activity of the immune system, while PCR is a better tool for screening and primary detection of disease. As for the availability of such a precise diagnostic tool, its importance is determined by the fact that opisthorchiasis is frequently asymptomatic. Moreover, it has been proved that one of the most severe complications of opisthorchiasis is cholangiocarcinoma, malignant tumor of bile ducts, which suggests that the population of endemic foci of opisthorchiasis should be examined for liver fluke infection on a regular basis and correspondingly treated when the disease is detected.

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